

# Chicken embryonic spinal cord electroporation

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## Organisation

**Name of the organisation** Université Catholique de Louvain (UCL)

**Department** Louvain Institute of Biomolecular Science and Technology (LIBST)

**Specific Research Group or Service** Animal Molecular and Cellular Biology (AMCB)

**Country** Belgium

## SCOPE OF THE METHOD

<b>The Method relates to</b>	Human health
<b>The Method is situated in</b>	Basic Research, Education and training
<b>Type of method</b>	In vivo
<b>Used species</b>	Chicken ( <i>Gallus gallus domesticus</i> )
<b>Targeted organ system or type of research</b>	Central and peripheral nervous system

## DESCRIPTION

### Method keywords

Embryonic spinal cord

Electroporation

In ovo experiment

Expression vector

RNA interference

Gene overexpression  
Gene downregulation  
chicken  
chicken embryo

### **Scientific area keywords**

Development  
neurobiology  
Developmental neurobiology  
Spinal cord  
CNS  
Neurogenesis  
Neuronal differentiation  
Neuronal migration  
Gene expression  
Embryo

### **Method description**

The goal of chicken embryonic spinal cord electroporation is to increase or to reduce expression levels of genes of interest in the developing spinal cord, and to assess the phenotypic consequences of these alterations on neuronal differentiation or migration. Fertilized eggs stored at 14°C are incubated for ~60 hours at 38°C to obtain embryos at the expected developmental stage. Plasmid DNA or siRNA is injected in the lumen of the neural tube at Hamburger-Hamilton stages ~10 to ~18. Later stages cannot be injected due to the rotation of the embryo. Nucleic acids are internalized in neural progenitors and their progeny on one side of the neural tube using whole-embryo electroporation. The contra-lateral side can be used as a perfect matching control. Instead of the spinal cord, hindbrain, midbrain, or forebrain can also be targeted by adapting the position of the electroporation electrodes. Specific cell populations can be targeted by using a cell-specific promoter. Signaling pathway reporter constructs or labelling systems targeting neurites or synapses can also be (co-)electroporated. Development can be continued for 1 to 5 days depending on the developmental stage to be analyzed.

## **Lab equipment**

- Egg storage cabinet (wine cooler) Haier
- Eppendorf FemtoJet injector
- Harvard Apparatus BTX ECM830 power source + electrodes
- FIEM egg incubators

## **Method status**

History of use

Internally validated

Published in peer reviewed journal

## **PROS, CONS & FUTURE POTENTIAL**

### **Advantages**

- cheap
- fast
- versatile regarding gene alterations, labelings, or reporter activity that can be obtained
- adapted for screening a reasonable number of candidate genes for a particular process

### **Challenges**

- requires some initial training and skill
- limited time-window for the injection/electroporation (~HH10 to ~HH18)
- transient activity of the injected constructs
- limited duration of the post-electroporation period of time
- variability from one embryo to another (can be reduced with practice)

## **REFERENCES, ASSOCIATED DOCUMENTS AND OTHER INFORMATION**

### **References**

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### **Associated documents**

[Chicken embryo electroporation - NEDI lab \(UCLouvain\).pdf](#)

[Fixation procedure for chicken embryos.pdf](#)

### **Links**

[Short movie of chicken embryonic spinal cord electroporation](#)

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