

# Assaying Cellular Viability Using the Neutral Red Uptake Assay

*Commonly used acronym: NRU*

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## SCOPE OF THE METHOD

<b>Alternative method relates to</b>	Human health
<b>Alternative method is situated in</b>	Basic Research, Education and training, Regulatory use - Routine production
<b>Type of alternative method</b>	In vitro - Ex vivo
<b>This method makes use of</b>	Human derived cells / tissues / organs
<b>Specify the type of cells/tissues/organs</b>	Hepatic cell lines such as HepG2, HepaRG. Other cell lines also possible, e.g. 3T3 mouse fibroblasts.

## DESCRIPTION

### Method keywords

cellviability

toxicity

acute toxicity

neutral red uptake

HepG2

### Scientific area keywords

in vitro toxicity

viability study

hepatic toxicity

basal toxicity

### **Method description**

The neutral red uptake assay is a cell viability assay that allows in vitro quantification of xenobiotic-induced cytotoxicity. The assay relies on the ability of living cells to incorporate and bind neutral red, a weak cationic dye, in lysosomes. As such, cytotoxicity is expressed as a concentration-dependent reduction of the uptake of neutral red after exposure to the xenobiotic under investigation. The neutral red uptake assay is mainly used for hazard assessment in in vitro toxicology applications.

### **Lab equipment**

Incubator ( $37 \pm 1$  °C,  $90 \pm 5\%$  humidity,  $5.0 \pm 1\%$  CO<sub>2</sub>/air)

Laminar flow / clean bench / cabinet (standard: "biological hazard")

Water bath ( $37 \pm 1$  °C)

Inverse-phase contrast microscope

Laboratory balance

96-Well plate spectrophotometer (i.e., plate reader) equipped with  $540 \pm 10$  nm filter

Shaker for microtiter plates

Cell counter or hemocytometer

Pipettes, pipettors (multichannel and single channel; multichannel repeater pipette)

96-Well flat-bottom tissue culture microtiter plates

Multichannel reagent reservoir

Vortex mixer

### **Method status**

Published in peer reviewed journal

Validated by an external party (e.g. OECD, EURL ECVAM,...)

### **PROS, CONS & FUTURE POTENTIAL**

#### **Advantages**

Fast

Accurate

Cheap

## Challenges

Relatively easy to perform

## REFERENCES, ASSOCIATED DOCUMENTS AND OTHER INFORMATION

### References

Ates, Gamze, Tamara Vanhaecke, Vera Rogiers, and Robim M. Rodrigues. "Assaying Cellular Viability Using the Neutral Red Uptake Assay." *Cell Viability Assays: Methods and Protocols* (2017): 19-26

### Associated documents

[NRU Book chapter.pdf](#)

## PARTNERS AND COLLABORATIONS

### Organisation

**Name of the organisation** Vrije Universiteit Brussel

**Department** Pharmaceutical and Pharmacological Sciences (FARM)

**Specific Research Group or Service** In Vitro Toxicology and Dermato-cosmetology

**Country** Belgium

*Coordinated by*



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