

# Fish embryo test for acute toxicity testing of chemicals

*Commonly used acronym: FET-test, OECD TG 236*  
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## SCOPE OF THE METHOD

<b>The Method relates to</b>	Environment
<b>The Method is situated in</b>	Regulatory use - Routine production
<b>Type of method</b>	In vitro - Ex vivo
<b>This method makes use of</b>	Other (e.g. bacteria): embryo of zebrafish, up to 4 days post fertilization

## DESCRIPTION

### Method keywords

Fish embryo

Fertilised zebrafish eggs

Somite formation

Fertilisation rate

Reproducibility

Lethality

Coagulation

LC50Fish embryo

Fertilised zebrafish eggs

LC50

### **Scientific area keywords**

Chemical testing

Ecotoxicity Acute toxicity

In vitro toxicologyChemical testing

EcotoxicityAcute toxicity

in vitro toxicology

### **Method description**

The method has been fully described in OECD TG 236 and is intended to determine the acute or lethal toxicity of chemicals on embryonic stages of fish (*Danio rerio*). Newly fertilized zebrafish eggs are exposed to the test chemical for a period of 96 hrs. Every 24 hrs. Twenty embryos (one embryo per well) are exposed to the chemical tested at each concentration level. The test includes five increasing concentrations of the chemical tested and a control. Every 24 hours, four apical observations are recorded as indicators of lethality: (i) coagulation of fertilised eggs, (ii) lack of somite formation, (iii) lack of detachment of the tail-bud from the yolk sac, and (iv) lack of heartbeat. At the end of the exposure period, acute toxicity is determined based on a positive outcome in any of the four apical observations recorded, and the LC50 is calculated. The test report also includes a number of other important information elements related to the conduct of the test, in particular: the concentration of dissolved oxygen, pH, total hardness, temperature and conductivity of solutions, measured concentrations of the chemical tested, and whether the validity criteria of the test were met.

### **Lab equipment**

- Fish tanks made of chemically inert material and of a suitable capacity;
- Inverted microscope or binocular with a capacity of at least 80-fold magnification;
- Test chambers (glass or polystyrene);
- Incubator or air-conditioned room with controlled temperature;
- Spawn trap.

### **Method status**

Published in peer reviewed journal

Validated by an external party (e.g. OECD, EURL ECVAM,...)

### **PROS, CONS & FUTURE POTENTIAL**

#### **Advantages**

- Fast and cost-effective;
- The FET-test has been successfully applied to a wide range of substances exhibiting diverse modes of action, solubility, volatility, and hydrophobicity.

#### **Challenges**

- If the method is used for the testing of a mixture, its composition should, as far as possible, be characterized. Before use for regulatory testing of a mixture, it should be considered whether it will provide acceptable results for the intended regulatory purpose.
- The metabolic capabilities of fish embryos are not always similar to that of juvenile or adult fish. For instance, the protoxicant allyl alcohol has been missed in the FET.
- For substances with a molecular weight  $\geq 3$ kDa, a very bulky molecular structure, and substances causing delayed hatch which might preclude or reduce the post-hatch exposure, embryos are not expected to be sensitive because of limited bioavailability of the substance, and other toxicity tests might be more appropriate.
- The use of solvents should be avoided, but may be required in some cases in order

to produce a suitably concentrated stock solution. Where a solvent is used to assist in stock solution preparation, its final concentration should not exceed 100 µl/L and should be the same in all test vessels. When a solvent is used, an additional solvent control is required.

For more information see OECD TG nr. 236

### **Future & Other applications**

The test can also be used for the selection and ranking of candidate chemicals during the development of new chemicals and products and in toxicology research.

## **REFERENCES, ASSOCIATED DOCUMENTS AND OTHER INFORMATION**

### **References**

Busquet F, Strecker R, Rawlings JM, Belanger SE, Braunbeck T, Carr GJ, Cenijn P, Fochtman P, Gourmelon A, Hübler N, Kleensang A, Knöbel M, Kussatz C, Legler J, Lillicrap A, Martínez-Jerónimo F, Polleichtner C, Rzodeczko H, Salinas E, Schneider KE, Scholz S, van den Brandhof EJ, van der Ven LT, Walter-Rohde S, Weigt S, Witters H, Halder M. (2014). OECD validation study to assess intra- and inter-laboratory reproducibility of the zebrafish embryo toxicity test for acute aquatic toxicity testing. Regul Toxicol Pharmacol. 69 (3), 496-511

### **Associated documents**

[Busquet Witters et al 2014\\_Regul Tox Pharmacol.pdf](#)  
[OECD Test No. 236.pdf](#)

### **Links**

[OECD guideline n°236](#)

## PARTNERS AND COLLABORATIONS

### Organisation

**Name of the organisation** VITO

**Department** Health

**Country** Belgium

**Geographical Area** Flemish Region

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